# Statins reduce T cell inflammatory and pathogenic responses by inducing Kruppel-like 

 factor 2 expressionDe-xiu Bu ${ }^{1}$, Margarite Tarrio ${ }^{1}$, Nir Grabie ${ }^{1}$, Yuzhi Zhang ${ }^{1}$, Hiroyuki Yamazaki ${ }^{2}$, George Stavrakis ${ }^{1}$, Elena Maganto-Garcia ${ }^{1}$, Zachary Pepper-Cunningham ${ }^{1}$, Petr Jarolim ${ }^{1}$, Masanori Aikawa ${ }^{2}$, Guillermo García-Cardeña ${ }^{1}$, Andrew H Lichtman ${ }^{1}$.<br>${ }^{1}$ Department of Pathology, Brigham and Women's Hospital, Boston, Massachusetts, USA.<br>${ }^{2}$ Cardiovascular Division, Department of Medicine, Brigham and Women's Hospital, Boston, Massachusetts, USA.

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Nonstandard abbreviations used: KLF2, kruppel-like factor 2; S1PR1, sphingosine 1 phosphate receptor 1; PHA, phytohemagglutinin; IP-10, IFN- $\gamma$ inducible protein 10;GGPP, geranygeranyl pyrophosphate;FPP, farnesyl pyrophosphate; GGTI, geranyl geranyl transferase inhibitor;FTI, farnesyl transferase inhibitor.

Conflict of interest: The authors have declared that no conflict of interest exists.

| Gene | Forward Primer | Reverse Primer |
| :---: | :---: | :---: |
| MOUSE |  |  |
| $\beta$-actin | 5'-TCC TTC GTT GCC GGT CCA-3' | 5'-ACC AGC GCA GCG ATA TCG TC-3' |
| Klf2 | 5'-ACA GAC TGC TAT TTA TTG GAC CTT AG-3' | 5'-CAG AAC TGG TGG CAG AGT CAT TT-3' |
| Ccr 7 | 5'-CCA GAC CGT GGC CAA TTT CAA CAT-3' | 5'-ACA AGA AAG GGT TGA CAC AGC AGC-3' |
| Ccr 5 | 5'-ACT GCT GCC TAA ACC CTG TCA TCT-3' | 5'-TTC ATG TTC TCC TGT GGA TCG GGT-3' |
| Sell (Cd62l) | 5'-CAT TCC TGT AGC CGT CAT GG-3' | 5'-AGG AGG AGC TGT TGG TCA TG-3' |
| Ifng | 5'-AAC GCT ACA CAC TGC ATC TTG G-3' | 5'-GCC GTG GCA GTA ACA GCC-3' |
| Slpr 1 | 5'-GTG TAG ACC CAG AGT CCT GCG-3' | 5'-AGC TTT TCC TTG GCT GGA GAG-3' |
| Vcam1 | 5'-CCA AAT CCA CGC TTG TGT TGA-3' | 5'-GGA ATG AGT AGA CCT CCA CCT-3' |
| Cxcl10 | 5'-GCC GTC ATT TTC TGC CTC A-3' | 5'-CGT CCT TGC GAG AGG GAT C-3' |
| Ccl5 | 5 ${ }^{\prime}$ CAA GTG CTC CAA TCT TGC AGT C-3' | 5'-TTC TCT GGG TTG GCA CAC AC-3' |
| HUMAN |  |  |
| $\beta$-actin | 5'-GAG CTA CGA GCT GCC TGA CG-3' | 5 ${ }^{\prime}$-GTA GTT TCG TGG ATG CCA CAG GAC T-3' |
| KLF2 | 5'-CTT TCG CCA GCC CGT GCC GCG-3' | 5'-AAG TCC AGC ACG CTG TTG AGG-3' |
| IFNG | 5'-ATA TTT TAA TGC AGG TCA TTC AGA TGT AG-3' | 5'-TGA AGT AAA AGG AGA CAA TTT GGC T-3' |
| CD59 | 5'-ATG CGT GTC TCA TTA C-3' | 5'-TTC TCT GAT AAG GAT GTC-3' |

Supplemental Table 1: Oligonucleotide primers used for qRT-PCR


Supplemental Figure 1. Dose response of pitavastatin induction of Klf2 mRNA in effector mouse T cells. In vitro-generated effector OT-1 cells were treated with the indicated concentrations of pitavastatin for 18 h before RNA isolation and qRT-PCR analysis of Klf2.


Supplemental Figure 2. Statins increase mouse CD4 ${ }^{+}$T cell Klf2 mRNA expression. Naïve OT-1I (CD4 ${ }^{+}$) T cells were treated with the vehicle only (control), or $10 \mu \mathrm{M}$ simvastatin (Simva ) for 18 h , then stimulated with $\alpha \mathrm{CD} 3$ for 6 h before RNA isolation and qRT-PCR analysis of $K l f 2$ (A). In vitro-generated effector OT-II cells were treated with the vehicle (control) or simvastatin for 18 h , before RNA isolation and qRT-PCR analysis of $K l f 2$. The data are the mean $\pm$ s.d. of two experiments. C57BL/6 mice were injected i.p. with $20 \mathrm{mg} / \mathrm{kg}$ lovastatin or DMSO vehicle control for 3 consecutive days. Splenic $\mathrm{CD}^{+}$T cells were purified and cultured with or without $\alpha$-CD3 for 6 h, RNA was isolated and qRT-PCR analysis was performed for $K l f 2$ expression (C), Data in $C$ are the mean $\pm$ S.E.M. from 3 experiments.


Supplemental Figure 3. In vivo statin treatment reduces pathogenicity of $\mathbf{T}$ cells. cMy-mOva mice were fed pitavastatin ( $30 \mathrm{mg} / \mathrm{kg}$ ) or vehicle (control) by gavage twice per day for 8 days consecutive days. On the third day of statin treatment, $5 \times 10^{4}$ effector OT- 1 cells were adoptively transferred into the mice. The day following the last pitavastatin treatment, the mice were sacrificed, and serum was sampled for determination of troponin I levels. Data are the mean $\pm$ S.E.M. of samples from 4 or 5 mice in each group.

